THE GENERA OF PARASITIC HYMENOPTERA IN THE PHILIPPINES, Part 1*

By Clare R. Baltazar

BUREAU OF PLANT INDUSTRY, MANILA

INTRODUCTION

Students in biological control work in the Philippines need to know the parasitic groups of Hymenoptera found in this area. The keys to genera presented here are designed to help identify adult parasites encountered in the field or reared from various host insects.

This paper will be published in several parts containing five sections: I. Megalyridae, Stephanidae, Trigonalidae, Gasteruptiidae and Evaniidae, II. Braconidae, III. Ichneumonidae, IV. Chalcidoidea and V. Proctotrupoidea. Part 1 includes the first two sections; the other sections will follow in the order given above. In the Philippines the group with the largest number of genera is the family Ichneumonidae followed by the Chalcidoidea, Braconidae, and the Proctotrupoidea. The rest are small families and have only a few members.

The genera included are those reported in my catalogue of Philippine Hymenoptera (in press) and new records of genera discovered after studying Philippine specimens in various collections and museums like Baker's collection deposited in the United States National Museum, Washington, D. C.; Townes' collection in the University of Michigan, Ann Arbor; Chicago Museum of Natural History, Chicago; British Museum of Natural History, London; Hope Museum in Oxford University, Oxford; Naturhistoriska Riksmuseet, Stockholm; Museum National d'Histoire Naturelle, Paris; Museum d'Histoire Naturelle, Geneva; Bishop Museum, Honolulu; Bureau of Plant Industry, Manila; and collection of the College of Agriculture, University of the Philippines, Los Baños.

The grouping into superfamilies was based on the Hymenoptera Synoptic Catalog of the USDA, published as Monograph No. 2 (1951) by Muesebeck and others, and its supplement (1958) by Krombein and others. Richards (1956) differed in recognizing superfamilies Evanioidea and Trigonaloidea.

Synopsis of the Parasitic Hymenoptera in the Philippines

Superfamily Ichneumonoidea

Family Megalyridae..... The Megalyrids

^{*} This research was conducted in the United States and Europe, 1957-58, with the financial assistance of the John Simon Guggenheim Memorial Foundation, New York, to which the writer wishes to express her sincere appreciation.

Family Stephanidae The Stephanids
Family Braconidae The Braconids
Family Ichneumonidae The Ichneumon-flies
Superfamily Chalcidoidea The Chalcid flies
Family Trichogrammatidae The Trichogrammatids
Family Mymaridae The Fairy-flies or Mymarids
Family Eulophidae The Eulophids
Family Eupelmidae The Eupelmids
Family Encyrtidae The Encyrtids
Family Agaonidae The Fig insects
Family Torymidae The Torymids
Family Chalcididae The Chalcidids
Family Eucharitidae The Eucharitids
Family Pteromalidae The Pteromalids
Family Eurytomidae The Eurytomids
Family Perilampidae The Perilampids
Superfamily Cynipoidea The Gall-flies or Gall wasps
Family Liopteridae The Liopterids
Family Cynipidae The Cynipids
Superfamily Proctotrupoidea (=Serphoidea)
Family Evaniidae The Ensign-flies
Family Gasteruptiidae The Gasteruptiids
Family Proctotrupidae The Proctotrupids
Family Ceraphronidae The Ceraphronids
Family Diapriidae The Diapriids
Family Scelionidae The Scelionids
Family Platygasteridae The Platygasterids
Superfamily Bethyloidea
Family Chrysididae The Cuckoo wasps
Family Loboscelidiidae The Loboscelidiids
Family Bethylidae The Bethylids
Family Dryinidae The Dryinids
Family Trigonalidae

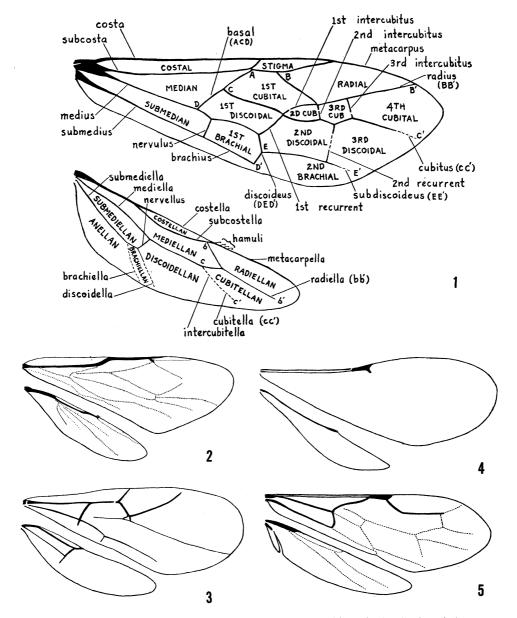
The Cynipoidea found in the Philippines has been incorporated in Weld's monograph of the Cynipoidea (1952), therefore it will not be discussed here.

A number of Hymenoptera which are classed by many authors as Aculeata are exclusively parasitic. The principal examples are the families listed under Bethyloidea. These and the scattered genera of parasitic Aculeata are not covered by this paper.

The genus *Loboscelidia* Westwood which was placed by Ashmead in the Cynipoidea and in the family Diapriidae (Proctotrupoidea) by Dalla Torre, Kieffer and Fouts is now removed to the superfamily Bethyloidea on account of the presence of an anal lobe in the hind wing. Being unique in the characters mentioned in the key, couplet 14, the family Loboscelidiidae was erected for the genus *Loboscelidia* by Maa and Yoshimoto (1961).

The following key was adopted from Brues, Melander and Carpenter's classification of

insects (1954, p. 626) and Townes' key to the families of the commoner Clistogastra appearing in Comstock's Introduction to Entomology (1948). This key starts out with the suborder Apocrita or Clistogastra, or Hymenoptera with the abdomen deeply constricted



Figs. 1-5. 1, wing venation of *Poecilogonalos* sp. (Trigonalidae) (Veins in dotted lines were added to indicate the position of the cubitellan and brachiellan cells.); 2, wings of *Brachymeria euploeae* Westwood (Chalcididae); 3, wings of *Saphonecrus areolatus* Weld (Cynipidae); 4, wings of *Aparamesisu microtomus* Kieffer (Diapriidae); 5, wings of bethylid (Bethylidae).

between the propodeum and the gaster. In this suborder cenchri are never present; fore tibia has only one apical spur; fore wing has no closed anal cell.

The system of wing venation used is that by Rohwer and Gahan (1916) but in the family Ichneumonidae "areolet" was substituted for the "second cubital cell." The wing illustrations are intended to show only venation without indicating infuscated areas or the arrangement of setae.

KEY TO SUPERFAMILIES AND FAMILIES OF PHILIPPINE PARASITIC HYMENOPTERA

1.	Wings present
	Wings absent16
2(1).	Antenna with more than 13 segments or hind wing without closed cells (figs.
	16-18); legs usually with 2 trochanters (fig. 6); ovipositor elongate or
	partly exposed and issuing before tip of abdomen (figs. 6-9); fore wing with
	or without costal cellParasitic Hymenoptera or Terebrantia, 3
	Antenna with 13 or fewer segments and hind wing with closed cells; legs
	each with a single trochanter; ovipositor (sting) entirely hidden and issu-
	ing from tip of abdomen; fore wing always with costal cell (fig. 1)
	Stinging Hymenoptera or Aculeata
3(2).	Costal and subcostal veins (Sc & R) separated in fore wing, enclosing nar-
5(2).	row costal cell (figs. 11–19); sternites sclerotized
	Costal and subcostal veins confluent in fore wing, costal cell absent or very
	narrow (figs. 29–73); sternites membranous or sclerotized
4 (2)	Mesoscutum with sharp median groove; abdomen elongate oval. (Outer orbit
4 (3).	of eye encircled by carina (fig. 6); small species, 3–8 mm long excluding
	• • • • • • • •
	ovipositor, Genus <i>Ettchellsia</i>) Family Megalyridae
- 5.5	Mesoscutum without median groove; abdomen variable
5 (4).	Abdomen inserted near top of propodeum, far above hind coxae (figs. 8-10);
	antenna with 13 or 14 segments
	Abdomen inserted normally, low down and close to hind coxae (fig. 7); an-
	tenna with 14–30 segments or more
6 (5).	Hind wing with anal lobe (figs. 16-18); fore wing with radial cell short and
	ending before apex of wing or absent; prothorax short; abdomen orbicular,
	compressed, borne on pedicel (fig. 10)Family Evaniidae
	Hind wing without anal lobe; fore wing with radial cell reaching apex of
	wing (figs. 14 & 15); prothorax prolonged into a neck; abdomen elongate,
	gradually clavate (figs. 8 & 9) Family Gasteruptiidae
7 (5).	Fore wing with 2 or 3 closed cubital cells; hind wing with 2 closed cells
	(fig. 19); head large, quadrate; mandible broad, quadridentate (fig. 20);
	antenna with 14 segments to more than 20; mesopleurum with transverse
	sutureFamily Trigonalidae
	Fore wing with 1 closed cubital cell (figs. 11-13); hind wing without closed
	cell; head globose and tuberculate above (fig. 7); mandible not unusually
	large, bidentate; antenna setaceous, with 30 segments or more; hind femur
	swollen and toothed before apexFamily Stephanidae
8 (3).	Antenna with 17 or more segments, rarely as few as 15; hind corner of pro-

notum reaching tegula (figs. 6 & 21); venter membranous and with longitudinal fold in dried specimen Superfamily Ichneumonoidea, 9 (excl. Megalyridae & Stephanidae) Antenna with 16 or fewer segments; hind corner of pronotum distant or not reaching tegula; venter sclerotized......10 9(8). Fore wing with 2 recurrent veins (as in fig. 6); all abdominal segments freely movable, except in very rare cases...... Family Ichneumonidae Fore wing with 1 or without any recurrent vein (figs. 29-73); abdominal segments 2 and 3 immovably united except in Aphidiinae Family Braconidae Hind wing with anal lobe (fig. 5) Superfamily Bethyloidea, 13 11 (10). Hind corner of pronotum reaching tegula (as in figs. 6 & 21); prepectus present; mid tibia with 1 apical spur, rarely absent; fore wing with single vein that is usually forked at apex (fig. 2); antenna elbowed..... Hind corner of pronotum not reaching tegula; prepectus absent; mid tibia 12 (11). Abdomen compressed and usually with mid-dorsal keel, polished, covered mostly by single tergite; costal vein absent; marginal cell (2d R_1+R_2) large (fig. 3)..... Superfamily Cynipoidea Abdomen cylindrical or depressed; costal vein present or marginal cell small or absent (fig. 4).....Superfamily Proctotrupoidea (excl. Evaniidae & Gasteruptiidae) 13 (10). Body brilliantly metallic; abdomen usually with 3 exposed tergites, seldom 4; venter concave...... Family Chrysididae Body usually black, not greenish or bluish; abdomen with more than 4 ex-14 (13). Antennal ledge distinct; abdomen short; membranous lamina present on scape. neck region, femora and tibiae; dorsal cervical plate present behind ocellar area and touching front margin of pronotum......Family Loboscelidiidae 15 (14). Antenna 10-segmented; front tarsus of 9 usually chelate Family Dryinidae Antenna 12 to 13-segmented; front tarsus of \mathcal{P} simple...... Family Bethylidae 16(1). Antenna elbowed; hind corner of pronotum not reaching tegula; prepectus Antenna not elbowed; hind corner of pronotum reaching tegula; prepectus 17 (16). Sternites 2 and 3 membranous, with longitudinal fold in dried specimens; ovipositor exposed beyond tip of abdomen Family Ichneumonidae Sternites 2 and 3 sclerotized, without mid-longitudinal fold; ovipositor usual-18 (17). Front tarsus chelate; antenna 10-segmented Family Dryinidae Front tarsus normal 19 19 (18). Abdomen compressed and usually with mid-dorsal keel, covered mostly by Abdomen depressed or cylindrical; head elongate with antenna inserted close

to anterior end Family Bethylidae

I. MEGALYRIDAE, STEPHANIDAE, TRIGONALIDAE, GASTERUPTIIDAE and EVANIIDAE

The five families discussed here, although belonging in different superfamilies have something in common, that is, they have the costal and subcostal veins (Sc & R) in the fore wing separated thus enclosing a narrow costal cell (figs. 11–19); also the sternites are sclerotized. The rest of the parasitic genera have the costal and subcostal veins confluent so there is no costal cell or the costal cell is very narrow.

In Richard's key to families of British Hymenoptera (1956), the families Evaniidae, Aulacidae, and Gasteruptiidae were grouped to constitute the superfamily Evanioidea or those with the gaster attached near the top of the propodeum. The superfamily Trigonaloidea was erected for the family Trigonalidae.

Synopsis of Genera

(Those marked with an asterisk are new records for the Philippines)

Family Megalyridae:

- 1. Ettchellsia Cameron
- Family Stephanidae:
 - 1. Megischus Brullé
 - 2. Parastephanellus Enderlein
 - 3. Foenatopus Smith

Family Trigonalidae:

- 1. Bakeronymus Rohwer
- 2. Trigonalys Westwood
- 3. *Poecilogonalos Schulz

Family Gasteruptiidae : Subfamily Aulacinae

Aulacus Jurine
*Aulacostethus Philippi

Subfamily Gasteruptiinae

Gasteruption Latreille

- 2. **Rhydinofoenus* Bradley
- Family Evaniidae:
 - 1. Evania Fabricius
 - 2. Prosevania Kieffer
 - 3. Parevania Kieffer
 - 4. Szepligetella Bradley

Family MEGALYRIDAE Fig. 6.

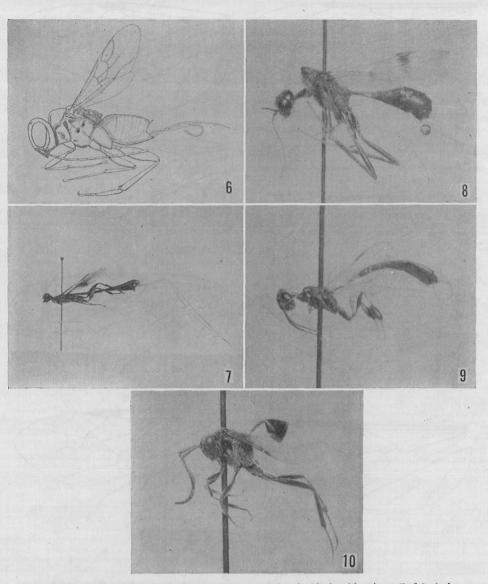
The family is mostly Indo-Australian in distribution and composed of large species with extremely long and prominent ovipositors. However, the species in the Philippines is small, measuring about 8 mm long including ovipositor. It belongs in the subfamily Dinapsinae. *Dinapsis* is reported from S. Africa.

The distinguishing characteristic of the family is the presence of a sharp median groove or linear furrow on the mesoscutum. The body is more or less cylindrical and the abdomen is elongate-oval.

In the Philippines, *Ettchellsia* is the only genus known in this family (Baltazar, 1962). The genus is distinct in having the outer orbit of the eye encircled by a carina (fig. 6).

Family STEPHANIDAE Figs. 7, 11, 12 & 13.

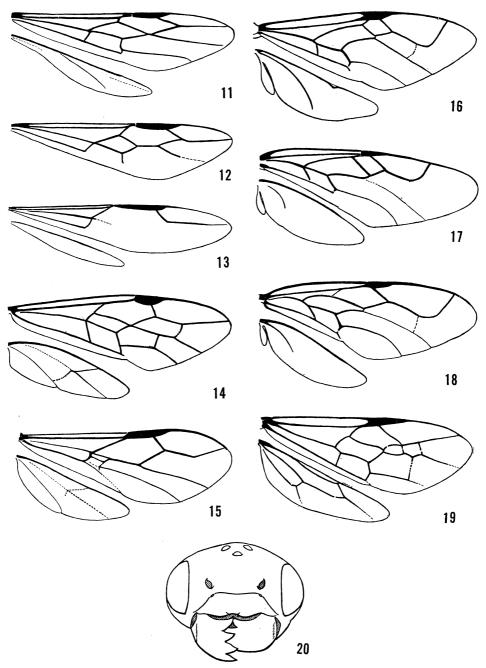
The stephanids are rather common in the tropics, often collected on dead tree trunks.



Figs. 6-10. 6, Ettchellsia philippinensis Baltazar (Megalyridae), side view; 7, Megischus coronator Fabricius (Stephanidae), side view; 8, Aulacostethus sp. (Gasteruptiidae), side view; 9, Gasteruption sp. (Gasteruptiidae), side view; 10, Prosevania sp. (Evaniidae), side view.

No host records are known in the family except for one record from Agrilus kalshoveni in Java parasitized by *Diastephanus leucosticus*. The species are presumed to be parasites of wood-boring coleopterous larvae or solitary bees or wasps nesting in wood.

The spherical head with a crown of teeth around the median ocellus is on a long neck. The antenna is setaceous, with 30 or more segments and arises just above the mouth. The abdomen is stalked, hind coxa elongate, hind femur swollen, and ovipositor long (fig.



Figs. 11-20. 11, Megischus sp. (Stephanidae), fore and hind wings; 12, Parastephanellus sp. (Stephanidae), fore wing; 13, Foenatopus sp. (Stephanidae), fore and hind wings; 14, Aulacostethus sp. (Gasteruptiidae), fore and hind wings; 15, Gasteruption sp. (Gasteruptiidae), fore and hind wings; 16, Evania appendigaster L. (Evaniidae), fore and hind wings; 17, Prosevania sp. (Evaniidae), fore and hind wings; 18, Parevania nitida Kieffer (Evaniidae), fore and hind wings; 19, Poecilogonalos sp. (Trigonalidae), fore and hind wings; 20, same, front view of head.

Baltazar: Parasitic Hymenoptera in the Philippines

7). The mandibles are bidentate.

KEY TO GENERA OF PHILIPPINE STEPHANIDAE

- Intercubital cell 3 and discoidal cell 1 separated by a stalk or cubitus vein with 3 abscissae; recurrent vein 1 not broken (fig. 12) Parastephanellus Intercubital cell 3 and discoidal cell 1 adjacent to each other or cubitus vein with 2 abscissae; recurrent vein 1 broken (fig. 11)...... Megischus

Family TRIGONALIDAE Figs. 19 & 20.

This is a small family, with only three genera recorded in the Philippines. No host records are known for the Philippine species but members of the family Trigonalidae are reported to be parasites, usually secondary parasites. Their hosts are likely to be the Vespidae, dipterous and hymenopterous parasites of caterpillars.

The taxonomic position of the trigonalids is uncertain. Townes (1956), in his paper on the nearctic species of trigonalid wasps, placed it with the Chrysidoidea (incl. the Bethyloidea) although he mentioned relationships to the Aculeata. He briefly distinguished the trigonalids from other families of Apocrita by the following: "Flagellum with 14 to more than 20 segments; costal cell of fore wing present; hind wing with distinct venation and two closed cells; anal lobe presented by a small vestige; and legs usually with two but sometimes with one trochanter each."

The mandibles are broad with four apical teeth on the right (fig. 20) and three on the left, mesopleurum with a transverse section, claws apically bifid, hind basitarsus equal to tarsal segments 2–5 united.

The occurrence of the genus *Poecilogonalos* in the Philippines is recorded here for the first time.

KEY TO GENERA OF PHILIPPINE TRIGONALIDAE

Family GASTERUPTIIDAE

The family is composed of two subfamilies, Aulacinae and Gasteruptiinae, although

some would treat these two groups as distinct families. Townes (1950) listed three characters that would readily separate the Gasteruptiidae from the other Hymenoptera: "1, abdomen attached to the thorax (alitrunk) high, far above the attachment of the hind coxae; 2, hind wing without closed cells or with a single closed cell; 3, first abdominal segment not set off from the rest of the abdomen by a conspicuous articulation. Of these, the first character is shared only with the Evaniidae and a few Braconidae and Ichneumonidae (most Labenini). The second character differentiates the Gasteruptiidae from the Braconidae and Ichneumonidae, and the third differentiates them from the Evaniidae."

Key to subfamilies of Gasteruptiidae

Subfamily AULACINAE Figs. 8 & 14.

The Aulacinae are reported to be parasites of wood-boring Coleoptera and Symphyta. In the Philippines there are no host records known and specimens in this group are not often encountered in collections, in fact, the genus *Aulacostethus* is a new record for the Philippines. Two genera in the subfamily may be differentiated as follows:

KEY TO GENERA OF AULACINAE

Subfamily GASTERUPTIINAE Figs. 9 & 15.

The gasteruptiines are parasitic on Sphecoidea and Apoidea nesting in wood or twigs, however, there are no host records in the Philippines. Some species may be caught around flowers. Two genera, *Gasteruption* and *Rhydinofoenus*, are differentiated in the following key. *Gasteruption* is mostly Holarctic in distribution unlike *Rhydinofoenus* which is a large genus and world-wide in distribution. The latter has not been reported from the Philippines before.

Baltazar: Parasitic Hymenoptera in the Philippines

KEY TO GENERA OF GASTERUPTIINAE

Family EVANIIDAE Figs. 10, 16, 17 & 18.

The evaniids commonly known as ensign-flies are parasitic in the egg capsules of Blattidae. This family has a large number of species in the tropics. Four genera are known in the Philippines.

The family is easily distinguished in having the gaster attached high or near the top of the propodeum by a cylindric and slightly arched petiole (fig. 10); the gaster is strongly compressed, circular, subcircular or subtriangular. The Evaniidae differs from other parasitic Hymenoptera in having a long anal lobe in the hind wing (figs. 16–18).

Key to genera of Philippine Evaniidae

II. BRACONIDAE Figs. 21–73.

The family Braconidae is one of the major groups of insect parasites and includes a

great number of species that are of considerable value in the biological control of insect pests. The family is almost entirely beneficial as primary parasites of Lepidoptera, Coleoptera, Diptera and Homoptera.

In the Philippines the braconid parasites and their corresponding hosts are listed in my catalogue of Philippine Hymenoptera (in press). Only 22 species have definite host records. Some common examples are *Apanteles* and *Chelonus* that are parasitic on many lepidopterous larvae, *Opius* on fruitfly maggots, and *Platyspathius* on powderpost beetle. Four parasites of the rice stem borers have been recorded, namely, *Bracon chinensis* (Szepligeti), *Tropobracon schoenobii* (Viereck), *Stenobracon nicevillei* (Bingham), and *Spathius fuscipennis* Ashmead. In 1955 attempts were made to introduce in the Philippines the polyembryonic parasite of the European corn borer, *Macrocentrus gifuensis* Ashmead from the United States, but were not successful. *Opius longicaudatus* Ashmead from the Philippines parasitizing *Dacus dorsalis* Hendel, was introduced to Hawaii and it successfully controls the fruitfly there. On the other hand *Opius fletcheri* Silvestri was introduced into the Philippines from Hawaii and it became established here and is successful against the cucurbit fruitfly, *Dacus cucurbitae* Coq.

The Braconidae is closely related to the Ichneumonidae and most species are easily distinguished from the latter by the absence of recurrent vein 2, the joint between abdominal segments 2 and 3 is inflexible except in Aphidiinae, and usually there is a vein crossing the large cell situated below the stigma.

At present the number of braconid species known in the Philippines is 197 in 63 genera. This paper includes 125 genera, 62 of which are considered new records for this area, with many more unidentified genera not included in the key.

I am indebted to Mr. C. F. W. Muesebeck of the U.S. National Museum, Washington, D. C., the foremost specialist in Braconidae, who checked the identifications of the specimens and made corrections on the manuscript. Many thanks are due Mr. G. E. J. Nixon of the British Museum of Natural History in London, who gave suggestions for the key; Dr. and Mrs. Townes who loaned their collections of the Philippine Braconidae for this study; and to the authorities of the U. S. National Museum who allowed me the use of the facilities in Washington, D. C., where most of this work was done.

Synoptic list of genera

(Those marked with an asterisk are new records for the Philippines)

Subfamily Aphidiinae:

- 1. **Ephedrus* Haliday
- 2. Diaeretus Foerster
- 3. * Monoctonus Haliday
- 4. * Aphidius Nees
- Subfamily Euphorinae:
 - 5. Aridelus Marshall
 - 6. Meteorus Haliday
 - 7. *Wesmaelia Foerster
 - 8. *Perilitus Nees
 - 9. *Euphoriana Gahan

- 10. **Euphorus* Nees
- 11. *Steblocera Westwood
- 12. * Microctonus Wesmael
- 13. *Syntretus Foerster

Subfamily Macrocentrinae:

- 14. Macrocentrus Curtis
- Subfamily Helconinae*:
 - 15. *Zele Curtis
 - 16. *Baeacis Foerster
 - 17. * Diospilus Haliday

- . .
 - 18. *Helcon Nees
 - 19. *Helconidea Helcon
- 20. *Cenocoelius Westwood
- Subfamily Blacinae:
 - 21. Orgilus Haliday
 - 22. *Centistes Haliday
 - 23. *Eubadizon Nees
 - 24. *Blacus Nees
 - 25. Stantonia Ashmead
 - 26. *Triaspis Haliday
 - 27. *Urosigalphus Ashmead
- Subfamily Agathidinae:
 - 28. Mesocoelus Schulz
 - 29. *Isoptronotum Enderlein
 - 30. Cremnops Foerster
 - 31. Disophrys Foerster
 - 32. *Zelomorpha Ashmead
 - 33. Euagathis Szepligeti
 - 34. Laccagathis Watanabe
 - 35. Braunsia Kriechbaumer
 - 36. Agathis Latreille
 - 37. *Camptothlipsis Enderlein
 - 38. *Baeognatha Kokujev

Subfamily Microgasterinae:

- 39. * Mirax Haliday
- 40. Apanteles Foerster
- 41. Microplitis Foerster
- 42. Microgaster Latreille
- 43. Snellenius Westwood
- 44. Fornicia Brullé
- Subfamily Cardiochilinae:
 - 45. Cardiochiles Nees
 - 46. Laminitarsus Fullaway
- Subfamily Cheloninae:
 - 47. Phanerotoma Wesmael
 - 48. Phanerotomella Szepligeti
 - 49. Ascogaster Wesmael
 - 50. Megascogaster Baker
 - 51. Cubochelonus Baker
 - 52. Chelonus (Chelonus Jurine) Chelonus (Neochelonella Hincks)

Subfamily Alysiinae:

- 53. *Synaldis Foerster
- 54. *Dinotrema Foerster

- 55. * Aspilota Foerster
- 56. Alysia Latreille
- 57. *Idiasta Foerster
- 58. * Anarcha Foerster
- 59. *Cratospila Foerster
- 60. * Acrobela Foerster
- 61. *Phaenocarpa Foerster
- 62. Asobara Foerster
- Subfamily Dacnusinae*:
 - 63. *Symphya Foerster
 - 64. * Dacnusa Haliday
 - 65. *Coelinius Nees
- Subfamily Opiinae:
 - 66. Opius Wesmael
- Subfamily Braconinae:
 - 67. *Batotheca Enderlein
 - 68. Spinaria Brullé
 - 69. *Spinariella Szepligeti
 - 70. *Gastrotheca Guerin
 - 71. Aphrastobracon Ashmead
 - 72. Chaoilta Cameron
 - 73. *Atanycolus Foerster
 - 74. Stenobracon Szepligeti
 - 75. Gronaulax Cameron (=Neuraulax Roman)
 - 76. Euurobracon Ashmead
 - 77. *Vipio Latreille
 - 78. *Bathyaulax Szepligeti
 - 79. Iphiaulax Foerster
 - 80. Cratobracon Cameron
 - 81. Sigalphogastra Cameron
 - 82. *Stirobracon Cameron
 - 83. Tropobracon Cameron
 - 84. *Odontopygia Enderlein
 - 85. *Dioxybracon Granger
 - 86. * Myosoma Brullé
 - 87. Odontogaster Szepligeti
 - 88. Hemiglyptus Ashmead
 - 89. *Cratocnema Szepligeti
 - 90. *Philomacroploea Cameron
 - 91. Campvloneurus Szepligeti
 - 92. Bracon Fabricius

Subfamily Spathiinae:

93. *Pseudospathius Szepligeti

- 94. Paraspathius Nixon
- 95. Spathius Nees
- 96. Platyspathius Viereck

Subfamily Stephaniscinae*:

- 97. *Leptospathius Szepligeti
- 98. *Halycaea Cameron
- 99. *Doryctophasmus Enderlein

Subfamily Rogadinae:

- 100. * Yelicones Cameron
- 101. Dedanima Cameron (=Colastomion Baker)
- 102. Macrostomion Szepligeti (=Macrostomionella Baker)
- 103. Megarhogas Szepligeti
- 104. Clinocentrus Haliday
- 105. Rogas Nees
- 106. Conspinaria Schulz (=Paragyroneuron Baker)
- 107. Gyroneuronella Baker
- 108. Hemigyroneuron Baker

- 109. *Pelecystoma Wesmael
- 110. Rhogasella Baker
- 111. Pseudogyroneuron Baker
- 112. Aulosaphes Muesebeck
- 113. * Acanthormius Ashmead
- 114. * Pambolus Haliday
- 115. Hormius Nees
- Subfamily Doryctinae:
 - 116. *Odontobracon Cameron
 - 117. Euscelinus Westwood
 - 118. * Rhoptrocentrus Marshall
 - 119. Heterospilus Haliday
 - 120. Doryctes Haliday
 - 121. *Rhaconotus Ruthe
 - 122. Rhyssalus Haliday

Tribe Hecabolini:

- 123. *Aivalykus Nixon
- 124. Polystenus Foerster
- 125. Monolexis Foerster

The subfamilies of Braconidae

There are 17 subfamilies of Braconidae represented in the Philippines. Of these, three are new records for this country: subfamilies Helconinae, Dacnusinae and Stephaniscinae. The key to determine the subfamilies and tribes has been incorporated in the key to the genera. Below is a brief diagnosis or characteristic of each subfamily with its corresponding host selection.

Aphidiinae (fig. 34). All species are internal parasites of aphids. Plant lice with braconid cocoons inside have an inflated appearance and brownish color, often referred to as mummified aphids.

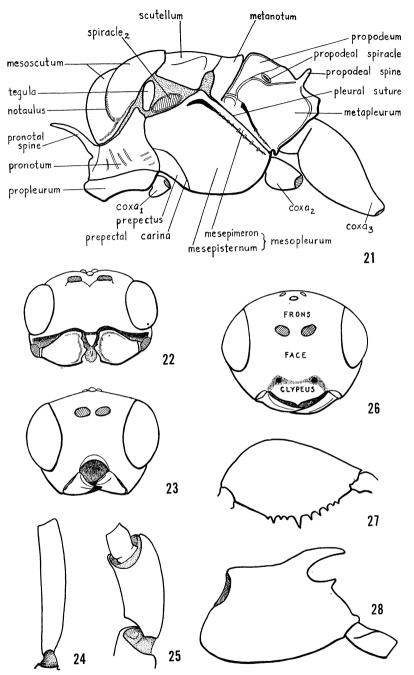
The species are usually small, not exceeding 4 or 5 mm. This subfamily is different from the other subfamilies in having the joint between tergites 2 and 3 flexible so the abdomen can fold toward the underside. The venation is generally reduced (fig. 34).

Euphorinae (figs. 24, 42–48). The great majority of species are internal parasites of adult Coleoptera. *Perilitus* is confined to Coccinellidae and Curculionidae whereas *Microctonus* attacks Curculionidae, Chrysomelidae and Tenebrionidae. According to Muesebeck (1936, 1951) *Aridelus* attacks nymphs and adults of Pentatomidae; *Euphorus* and *Euphoriana* are parasitic in Miridae. *Meteorus* is parasitic mostly on lepidopterous larvae and recorded also on wood-boring Coleoptera. No host records are known for the Philippine species.

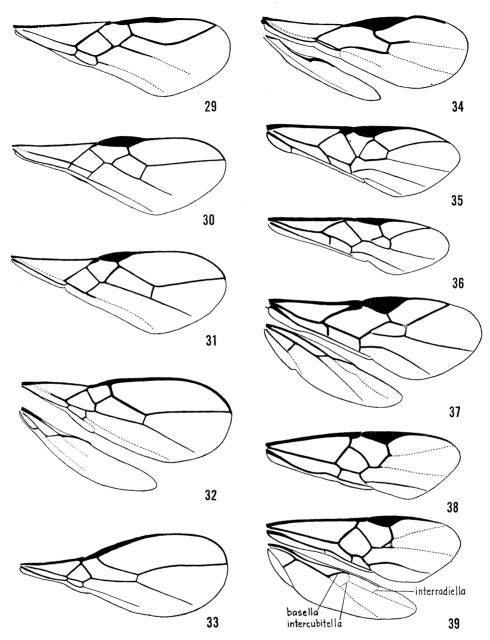
The euphorine species are usually small and black. In Muesebeck's (1936) treatise of the subfamily he characterized the Euphorinae in detail as follows:

"Head transverse to quadrate; mandibles bidentate, crossing at apices and fitting against clypeus; clypeus separated from face by an impression; anterior margin of clypeus

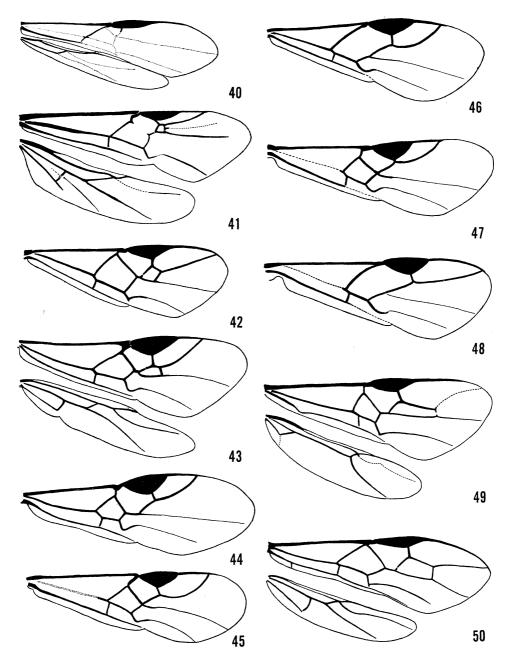
Baltazar: Parasitic Hymenoptera in the Philippines



Figs. 21-28. 21, Spinaria westwoodi flavipennis Roman (Braconinae), side view of thorax; 22, Idiasta sp. (Alysiinae), front view of head; 23, Bracon chinensis (Szepligeti) (Braconinae), front view of head; 24, Streblocera sp. (Euphorinae), scape; 25, Chaoilta intrudens Smith (Braconinae), scape; 26, Baeacis sp. (Helconinae), front view of head; 27, Euscelinus sarawacus Westwood (Doryctinae), hind femur; 28, Odontobracon sp. (Doryctinae), hind coxa.



Figs. 29-39. 29, Dacnusa sp. (Dacnusinae), fore wing; 30, Cratospila sp. (Alysiinae), fore wing; 31, Asobara bactrocerae Gahan (Alysiinae), fore wing; 32, Aspilota sp. (Alysiinae), fore and hind wings; 33, Synaldis sp. (Alysiinae), fore wing; 34, Aphidius sp. (Aphidiinae), fore and hind wings; 35, Phanerotoma ferruginea Baker (Cheloninae), fore wing; 36, Ascogaster philippinensis Baker (Cheloninae), fore wing; 37, Chelonus (C.) semihyalinus Ashmead (Cheloninae), fore and hind wings; 38, Apanteles sp. (Microgasterinae), fore wing; 39, Microgaster apo Wilkinson (Microgasterinae), fore and hind wings.



Figs. 40-50. 40, Mesocoelus philippinensis Muesebeck (Agathidinae), fore and hind wings; 41, Disophrys insignis Roman (Agathidinae), fore and hind wings; 42, Meteorus browni Ashmead (Euphorinae), fore wing; 43, Aridelus fumipennis Fouts (Euphorinae), fore and hind wings; 44, Perilitus sp. (Euphorinae), fore wing; 45, Wesmaelia sp. (Euphorinae), fore wing; 46, Microctonus sp. (Euphorinae), fore wing; 47, Euphorus sp. (Euphorinae), fore wing; 48, Syntretus sp. (Euphorinae), fore wing; 49, Cardiochiles sp. (Cardiochilinae), fore and hind wings; 50, Macrocentrus philippinensis Ashmead (Macrocentrinae) fore and hind wings.

subtruncate, more or less reflexed; maxillary palpi with 5 or 6 segments; labial palpi with 2, 3, or 4 segments; eyes prominent, usually bare or sparsely hairy, very rarely thickly hairy; temples and cheeks margined; occiput usually margined but with the carina often interrupted medially; transverse impression at base of scutellum broad and deep; prepectus margined; legs slender; calcaria of posterior tibia varying from very short to half as long as metatarsus; 2 or 3 cubital cells, very rarely only 1; radial cell ranging from very small and more or less lunate to very large, measured on wing margin in some specimens not more than one-fourth length of stigma, in others extending nearly to apex of wing; rarely, radial cell not defined, the radius lacking; first cubital and first discoidal cells either separated or confluent; recurrent vein very rarely absent; medius ranging from well developed to obsolete; first brachial cell open at apex; subdiscoideus not interstitial; submediellan cell large, rarely not defined. First abdominal tergite petiolate, the spiracles at or behind the middle; second and third tergites connate, large, usually but not always carinate at sides, usually overlapping on venter; apical margin of second tergite rarely distinct; ovipositor sheaths either concealed or prominent."

Macrocentrinae (fig. 50). These are internal parasites of lepidopterous larvae occurring in burrows or cavities in plant stems, fruit and seeds. The only known host record of *Macrocentrus* sp. in the Philippines was reported by Pierce (1928) on *Scirpophaga* sp. attacking sugar cane.

The species of *Macrocentrus* are yellowish brown or black, slender, with long legs and an ovipositor as long as the body. The subfamily is characterized by the absence of an occipital carina, the clypeus is convex, without an opening between clypeus and mandibles, cubital cell 2 is large and rectangular (fig. 50), and the spiracle on tergite 1 is before the middle.

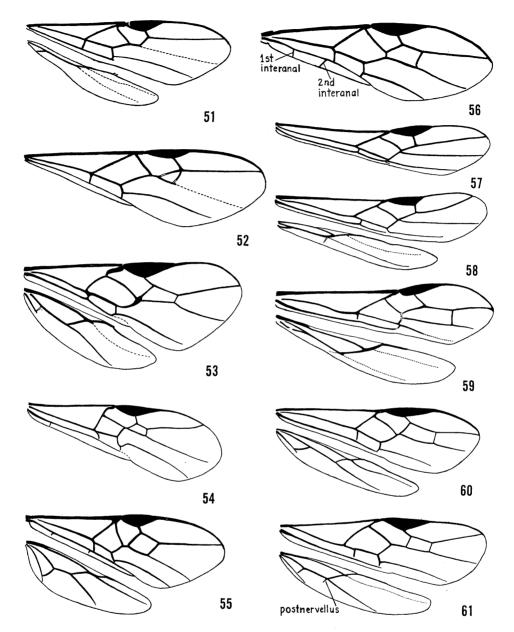
Helconinae (figs. 26, 54–56). These are parasites of wood-boring coleopterous larvae. No host records are known in the Philippines, in fact this is the first record of the occurrence of the subfamily here.

The Helconinae is composed of medium-sized species. The subfamily is characterized as follows: Clypeus convex with the space between clypeus and mandible narrow, occipital carina entire, fore wing with 3 cubital cells, anal cell of fore wing with 1 or 2 interanals, head usually conical or subcubical, and frontal depression more or less deep.

Blacinae (figs. 51 & 52). In this subfamily only two genera, Triaspis and Urosigalphus, are known to have host records. They are parasitic on coleopterous larvae of Bruchidae and occasionally Curculionidae. In the Philippines there are no host records for this subfamily. The 2 genera mentioned above are different from the rest of the Blacinae in that the abdomen is carapace-like, with only 2 sutures visible above as in the subfamily Cheloninae, but differ from the latter in having only 2 cubital cells instead of 3. Some authors place Triaspis and Urosigalphus in a separate family, Triaspinae.

The Blacinae has the clypeus convex, without an opening between the clypeus and mandibles, and the occipital carina is usually entire or dorsally interrupted as in *Orgilus*. The fore wing has 2 cubital cells except in *Stantonia* which has 3, and abscissa 2 of the radius is straight or almost straight and reaching the apex of wing.

Agathidinae (figs. 40 & 41). The hosts selected by the Agathidinae are lepidopterous larvae. So far as known there has been no host record in the Philippines. In Japan,



Figs. 51-61. 51, Orgilus ashmeadi Viereck (Blacinae), fore and hind wings; 52, Stantonia flava Ashmead (Blacinae), fore wing; 53, Opius fletcheri Silvestri (Opiinae), fore and hind wings; 54, Cenocoelius sp. (Helconinae), fore wing; 55, Baeacis sp. (Helconinae), fore and hind wings; 56, Helcon sp. (Helconinae), fore wing; 57, Aivalykus ecclectus Nixon (Hecabolini, Doryctinae), fore wing; 58, Polystenus ruficeps (Ashmead) (Hecabolini, Doryctinae), fore and hind wings; 59, Spathius apicalis Westwood (Spathiinae), fore and hind wings; 60, Pseudogyroneuron sp. (Rogadinae), fore and hind wings; 61, Rogas bicolor Baker (Rogadinae), fore and hind wings.

Mesocoelus philippinensis Muesebeck was recorded on Acrocercops transecta Meyrick by Yasumatsu and Kuroko (1957).

The subfamily may be recognized by the following set of characters: Clypeus convex, without an opening between clypeus and mandibles; cubital cell 2 very small; radial cell small, elongate and tapered toward apex of wing; radial vein strong; notaulus usually distinct; prepectal carina present; occipital carina absent, face sometimes much lengthened. *Mesocoelus* has the venation reduced, with only the nervulus, basal vein and abscissa 1 of radius distinct (fig. 40); hind coxa is elongate and as long as the hind femur.

Microgasterinae (figs. 38 & 39). The subfamily includes numerous species that are internal parasites of lepidopterous larvae. The genera of common occurrence are *Apanteles*, *Microgaster* and *Microplitis*.

The Microgasterinae has species with eyes usually hairy and the abdomen is very short so that it is surpassed by the hind femur. The distinguishing features are as follows: Cubital 2 small, triangular, and often confluent with cubital cell 3; radial cell large, radial vein weak or effaced towards apex of wing; notaulus absent; prepectal carina absent; clypeus not emarginate. *Snellenius* differs from the other Microgasterinae in having the notaulus deep and the prepectal carina present.

Cardiochilinae (fig. 49). Few host records are known for the subfamily but apparently they are parasites of lepidopterous larvae. No host records are known for the Philippine species in this subfamily.

The Cardiochilinae is easily distinguished by the strongly curved abscissa 3 of radius but the vein is usually weak. The legs are robust, the abdomen is sessile and short, and the ovipositor is curved.

Cheloninae (figs. 35-37). The species are predominantly solitary internal parasites of lepidopterous larvae. The parasite has the habit of ovipositing in the egg of the host and completing its larval development when the host larva is nearly mature.

The subfamily is easily recognized by the shape of the abdomen which is carapacelike or it looks like an inverted bath tub or a turtle shell. At most 2 sutures are visible on the abdomen dorsally. There are 3 cubital cells present on the fore wing, cell 2 being small and more or less triangular and definitely smaller than cell 3. As previously discussed in the subfamily Blacinae, *Triaspis* and *Urosigalphus* also have carapace-like abdomens but they possess only 2 cubital cells on the fore wing.

Alysiinae (figs. 22, 30-33) and Dacnusinae (fig. 29). These subfamilies have not been extensively studied. The common genera, like Alysia and Dacnusa, have been reported as internal parasites of fly maggots, Diptera. The only host record in the Philippines is Asobara bactrocerae Gahan on the nanka fruitfly, Dacus umbrosa Fabr.

This group of braconids is referred to as the Exodontes because of the peculiar mandibles (fig. 22) that have the teeth pointing outward instead of inward towards the mouth. The paddle-like mandibles are supposed to be used for digging in filth when searching for hosts.

Alysiinae has 3 cubital cells in the fore wing (except in *Synaldis*, fig. 33), cubital cell 1 being long because intercubitus 1 is lacking, and abscissa 1 of radius is about $4 \times$ as long as the intercubitus (figs. 30-32).

The Dacnusinae has 2 cubital cells in the fore wing, cubital cell 1 short since inter-

cubitus 1 is present, and abscissa 1 of the radius subequal in length to the intercubitus (fig. 29). The occurrence of this subfamily in the Philippines is recorded here for the first time.

Opiinae (fig. 53). These are parasites of Diptera. Several species of *Opius* in the Philippines have been reared from *Dacus* spp. and *Acidoxantha* sp.

The subfamily is composed of small insects, usually yellow or black. The members may be recognized as follows: Clypeus convex with opening between clypeus and mandible narrow, not a circular hole; occipital carina entire or interrupted dorsally; cell 2 is 4-sided; anal cell of fore wing without a cross-vein; head usually transverse, sometimes cubical. Some *Bracon* species may be mistaken for *Opius* or vice versa but the Braconinae have the opening between the clypeus and mandible circular and the clypeus hollowed out apically (fig. 23), also the occipital carina is wanting. There is a difference also in tergite 1, in that there is a V-shaped depression basally in *Bracon*, but not in *Opius*.

The 4 subfamilies discussed below belong to a group called Cyclostomi because of the circular opening formed between the concave clypeus and mandible. Members of the Cyclostomi usually parasitize lepidopterous larvae or beetle larvae living in tunnels or nests.

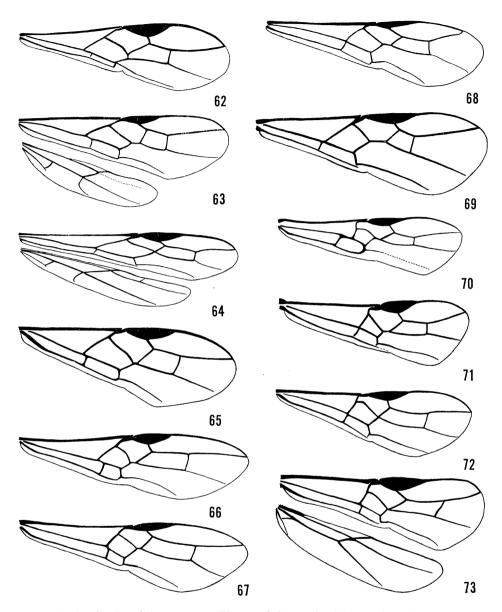
Braconinae (figs. 21, 23, 25, 65–73). The members of this subfamily are predominantly external parasites of caterpillars found in a cell, burrow or cocoon, some would attack coleopterous larvae, few are parasitic on sawflies and Cecidomyiidae. A species from the Philippines is predaceous rather than parasitic—Williams (1928) reported that Bracon lendicivorus (Cushman) develops at the expense of the cecidomyiid larvae, Asphotrophia fici Barnes, living in the fruit receptacles of Ficus nota. The common genus is Bracon (= Microbracon) which attacks a wide range of hosts. The first 3 parasites of rice stem borers mentioned in the introductory discussion of the family Braconidae belong in this subfamily.

This group is probably the largest in the subfamily and the most difficult to classify. The Braconinae is in bad need of revision. Among the Cyclostomi the Braconinae is different in having the occipital carina wanting, the head is usually transverse, the mesopleural furrow and prepectal carina are absent except in *Batotheca*, *Spinaria* and *Spinariella*, and the submediellan is small.

Spathiinae (fig. 59). Most of the species are external parasites of coleopterous borers, although a few are reported on lepidopterous borers. In the Philippines there are only 3 species with host records: *Platyspathius bisignatus* (Walker) and *P. dinoderi* (Gahan) parasitic on *Dinoderus minutus* F.; and *Spathius fuscipennis* Ashmead parasitic on *Chilo suppressalis* (Walker), one of the species of rice stem borers.

The Spathiinae is apparently differentiated from the other groups by a combination of characters: abdomen petiolate, tergite 1 not gradually widened apically except in *Platy-spathius*; epipleura not reaching middle of tergite; dividing suture between tergites 2 and 3 absent or faintly impressed in the Philippine genera; occipital carina present; fore wing with 3 cubital cells. Nixon's (1943) revision of the Spathiinae of the Old World should be consulted for the identification of the species.

Stephaniscinae (fig. 64). It is presumed that the species here are external parasites of wood-boring beetles since no host records have been reported for this group. The distribution of the Stephaniscinae seems to be in the Old World. Its occurrence in the Philip-



Figs. 62-73. 62, Euscelinus sarawacus Westwood (Doryctinae), fore wing; 63, Odontobracon sp. (Doryctinae), fore and hind wings; 64, Leptospathius sp. (Stephaniscinae), fore and hind wings; 65, Spinaria westwoodi flavipennis Roman (Braconinae), fore wing; 66, Euurobracon quadriceps apicalis Roman (Braconinae), fore wing; 67, Iphiaulax sp. (Braconinae), fore wing; 68, Stenobracon nicevillei (Bingham) (Braconinae), fore wing; 69, Tropobracon sp. (Braconinae), fore wing; 70, Aphrastobracon philippinensis Ashmead (Braconinae), fore wing; 71, Cratocnema sp. (Braconinae), fore wing; 72, Campyloneurus sp. (Braconinae), fore wing; 73, Bracon chinensis (Szepligeti) (Braconinae), fore and hind wings.

pines is reported here for the first time with four genera.

In the key presented in this paper, this subfamily will key out to the same couplet as Doryctinae, but may be differentiated as follows: Tergite 1 subpetiolate, widened at apex and about 2.5 or more times as long as apical width; petiole coarsely punctate or transversely striate but not longitudinally striate. These are medium to large species with long ovipositor.

Rogadinae (figs. 60–61). This subfamily is composed of large or medium-sized species parasitic on lepidopterous larvae; however, no host record is known in the Philippines except for *Aulosaphes psychidivorous* Muesebeck on a psychid larva on cacao. The parasitic larva develops and makes its cocoon inside its host.

In the rogadines the occipital carina, the mesopleural furrow and prepectal carina are present. It may be differentiated from the Doryctinae as follows, but sometimes with difficulty: ovipositor short, not longer than the greatest depth of abdomen; head usually transverse; fore tibia never with a row of spines on inner side; pronotal collar not swollen. Like the Braconinae and Doryctinae this group needs revision badly.

Doryctinae (figs. 27, 28, 57, 58, 62 & 63). Most of the species are external parasites of coleopterous larvae, a few might attack lepidopterous larvae. In the Philippines, Monolexis manilensis Ashmead was bred from a scolytid.

The Doryctinae is hard to define and often confused with the Braconinae but the latter lacks the occipital carina. This subfamily differs from the Rogadinae as follows: Ovipositor long, longer than the greatest depth of abdomen; head usually cubical; fore tibia usually armed with a row of short spines on inner side; pronotal collar usually swollen. A small subfamily, Stephaniscinae, will key out to the same diagnosis mentioned above but the Doryctinae has tergite 1 sessile, and if petiole is elongate the rest of the tergites are longitudinally striate.

KEY TO GENERA OF PHILIPPINE BRACONIDAE

1.	Mandibles widely separated, more or less flattened, teeth outward instead of
	inward, apices not touching nor overlapping when closed (fig. 22) 2
	Mandibles normal, apices touching or overlapping when closed (figs. 23 & 26)14
2(1).	Fore wing with 2 cubital cells
	Fore wing with 3 cubital cells (except in Synaldis)Alysiinae, 6
3 (2).	Cubital cell 1 long, intercubitus 1 wanting, abscissa 1 of radius about $4 \times$ as
	long as intercubitus (fig. 33) Synaldis
	Cubital cell 1 short, intercubitus 1 present, abscissa 1 of radius subequal in
	length to intercubitus (fig. 29) Dacnusinae, 4

DACNUSINAE

4 (3).	Abdomen	carapace-	like, on	ly suture	behind	tergite 1	present Symphya
	Abdomen	elongate,	sutures	between	tergites	present	

ALYSIINAE

(excl. Synaldis)

6 (2).	Stigma linear, joining costal vein imperceptibly; abscissa 2 of radius longer than intercubitus 1; radial cell more than 1/2 length of fore wing; cubital
	cell 2 is 5-sided (fig. 32)
	Stigma large, distinct from costal vein; abscissa 2 of radius variable in length
	in relation to intercubitus 1; radial cell less than or at most 1/2 length
	of fore wing; cubital cell 2 is 4-sided (figs. 30 & 31)
7(6).	Propodeal spiracle large and oval, rim somewhat raised Dinotrema
7(0).	Propodeal spiracle small, circular and puncture-like; propodeum carinate;
	sternaulus present
8(6).	Abscissa 2 of radius equal to or slightly shorter than intercubitus 1; radius
0(0).	arising beyond middle of stigma (fig. 30)
	Abscissa 2 of radius longer than intercubitus 1; radius arising from or near middle of stigma (fig. 31)
9(8).	Propodeal spiracle very large, propodeum with midlongitudinal carina Alysia
J (0):	Propodeal spiracle small, circular or puncture-like 10
10 (9).	Flagellar segment 1 shorter than 2; face wider than longAlysia
	Flagellar segment 1 longer than 2; face longer than wide11
11 (10).	Recurrent vein interstitial with intercubitus 1 Anarcha
	Recurrent vein antefurcal or entering cubital cell 1 (fig. 30) Cratospila
12 (8).	Flagellar segment 1 longer than 2; radius arising from the middle of stigma; postnervellus present
	Flagellar segment 1 shorter than 2; radius usually arising beyond the middle of stigma; postnervellus present or absent
12 (12)	Notaulus deeply impressed; postnervellus usually present Phaenocarpa
15(12).	Notaulus absent or impressed anteriorly; postnervellus absent (fig. 31) Asobara
14 (1).	Suture between tergites 2 and 3 membranous, thus abdomen can fold toward
14(1).	the underside; venation generally reduced (fig. 34); species small, not more
	than 4 or 5 mm
	Suture between tergites 2 and 3 rigid or erased; venation complete except in
	Mesocoelus; species small to large
	APHIDIINAE
15 (14).	Fore wing with 3 cubital cells Ephedrus
	Fore wing with less than 3 cubital cells 16
16 (15).	Fore wing with recurrent, cubitus and intercubitus veins effaced Diaeretus
	Fore wing with discocubital cell complete, recurrent vein effaced, at least a stub of cubitus always present
17 (16).	Abscissa 1 of radius nearly perpendicular to stigma; ovipositor and its sheaths
. ,	curved downward
	Abscissa 1 of radius oblique (fig. 34); ovipositor and its sheaths straight or
	curved upward
18 (14).	Clypeus convex, opening between clypeus and mandibles narrow or absent
	(fig. 26)
	Clypeus hollowed out apically, opening between clypeus and mandibles large

Baltazar: Parasitic Hymenoptera in the Philippines

	and more or less round (fig. 23)
19 (18).	Abdomen carapace-like, at most 2 sutures visible above
	Abdomen normal, with more than 2 sutures visible above
20 (19).	Fore wing with 2 cubital cells
	Fore wing with 3 cubital cells Cheloninae, 2.
21 (20).	Notaulus absent; head small, narrower than thorax; eyes hairy; abscissa 2
. ,	of radius weak; sutures on abdomen deep; scutellum dentate apically Fornicia
	Notaulus deep; head large, almost as wide as thorax; eyes bare; abscissa 2
	of radius distinct; abdomen with 2 weak transverse sutures present or absent;
	scutellum not dentate apically
22 (21).	Abdomen flattish, with 2 transverse sutures present, apical margin notched
()	and sometimes bidentate; antenna usually with more than 20 segments in \mathcal{J}
	Abdomen strongly convex, without transverse sutures, apically pointed in the
	Philippine species at hand; antenna seldom with more than 20 segments in
	a Urosigalphus
	CHELONINAE
23 (20).	Abdominal sutures distinct, 3 tergites visible above
	Abdominal sutures erased, tergites 1-3 forming a solid carapace
24 (23).	Radial vein with 3 abscissae; cubital cell 2 is 4-sided, large, often as large
	or larger than cubital cell 1 (fig. 35); eyes round Phanerotoma
	Radial vein with 2 abscissae; cubital cell 2 is 3-sided, small and elongate,
	smaller than cubital cell 1; eyes oblong Phanerotomella
25 (23).	Abscissa 1 of cubitus present, thus first cubital and first discoidal cells separate
	(fig. 36); eyes bare
	Abscissa 1 of cubitus absent, thus first cubital and first discoidal cells con-
	fluent (fig. 37); eyes hairy 27
26 (25).	Propodeal teeth present; abdomen rounded apically, or if pointed length at
	most 2.5 its greatest width; intercubitus 1 strongly arched toward stigma;
	abscissa 1 of radius equal to or longer than 2; nervulus far distad of basal
	vein (fig. 36)Ascogaster
	Propodeal teeth absent; abdomen pointed apically, length about 3.25 its
	greatest width; intercubitus 1 straight; nervulus shortly distad of basal vein
()	Megascogaster
27 (25).	Mesoscutum short, 0.7-0.8 as long as wide; head transverse from dorsal view;
	temple narrow; abdomen length less than 3× greatest width Chelonus, 28
	Mesoscutum long, 1.0-1.2 as long as wide; head subcubical from dorsal view;
	temple broad; abdomen length more than $3 \times$ its greatest width Cubochelonus
28 (27).	Female carapace with 2 apical teeth, $\vec{\sigma}$ carapace more or less acute apically;
	head wider than thorax; φ antenna with 16 segments Subgenus Neochelonella
	Female carapace without teeth, \mathcal{J} carapace rounded apically; head as wide
	or narrower than thorax; φ antenna with 16 or more segments
	Subgenus Chelonus
29 (19).	Venation reduced, only nervulus, basal vein and abscissa 1 of radius distinct
	(fig. 40); hind coxa elongate, as long as hind femur Mesocoelus

.

1962

•

Venation more or less complete; hind coxa shorter than hind femur (except 30 (29). Cubital cell 2 very small, triangular or squarish, sometimes open (figs. 38, 39 & 41); radial cell small, if large, radial vein weak towards wing apex; Cubital cell 2 large or medium, usually rectangular; radial cell large, if small, 31 (30). Radial cell large, radial vein weak or effaced towards apex of wing (figs. 38 & 39); notaulus absent or slightly impressed (except in Snellenius); prepectal carina absent (except in Snellenius)...... Microgasterinae, 32 Radial cell small, elongate and tapered towards apex of wing; radial vein strong; notaulus usually distinct (fig. 41); prepectal carina present..... Agathidinae, 36 MICROGASTERINAE (excl. Fornicia) 32 (31). Intercubitus 1 long, reaching stigma; radius not angled, obsolete except at extreme base; antenna with 14 segments Mirax Intercubitus 1 much shorter, not reaching stigma; radius with 2 abscissae, the first making an almost right angle with the second, the second usually indicated by a line of closely placed setae; antenna always with 18 segments. 33 33 (32). Areolet open, intercubitus 2 lacking (fig. 38) Apanteles 34 (33). Notaulus deep; mesoscutum raised and with midlongitudinal ridge; prepectal Notaulus absent or slightly impressed; mesoscutum evenly convex without 35 (34). Inner spur of hind tibia shorter than 1/2 length of hind basitarsus; hind coxa short, less than 1/2 as long as thorax; mesopleurum with distinct crenulate furrow...... Microplitis Inner spur of hind tibia usually more than 1/2 length of hind basitarsus; hind coxa long, at least as long as thorax; mesopleurum rarely with crenulate

furrow Microgaster

AGATHIDINAE

(excl. Mesocoelus)

36 (31). Fore and middle claws bifid	
Fore and middle claws simple, with or without a basal lobe	
37 (36). Claws pectinate at base; ovipositor long, about as long as abdomen; face	
elongate	
Claws with few long hairs at base; ovipositor short, not extending much be-	
yond tip of abdomen; face shorter	
38 (37). Notaulus absent; prepectal carina ending near top of mesopleurum; ovipositor	
sheath flat and wider than depth of ovipositor Isoptronotum	
Notaulus deep; prepectal carina ending at midheight of mesopleurum; ovi-	
positor sheath not wider than depth of ovipositor Cremnops	
20 (27) Submediation call large length reaching to shout 1/2 of mediation (for 11).	

39 (37). Submediellan cell large, length reaching to about 1/2 of mediellan (fig. 41);

	frontal carina bordering depression produced into a tooth near antennal
	base Disophrys
	Submediellan cell small, length at most reaching basal 1/3 of mediella;
	frontal carina absent, or if present not produced into a tooth near antennal
40 (00)	base
40 (39).	Frontal depression bordered by carinaZelomorpha
11 (00)	Frontal depression not bordered by carina Euagathis
41 (36).	Propodeum smooth; if carina present only a short midlongitudinal one and/
	or a basal transverse one; propodeal spiracle elongate and large
42 (41)	Propodeum rugose or mat; propodeal spiracle circular or oval, usually small 43 Frontal depression deep with a short medial carina above antennae; meso-
42 (41).	pleural furrow and notaulus absent
	Frontal depression shallow or absent, frons without a median carina; meso-
	pleural furrow present; notaulus present Braunsia
43 (41).	Areolet closed
	Areolet open
44 (43).	Propodeum and tergites 1 and 2 mat; clypeus about $2 \times$ as long as wide
	Camptothlipsis
	Propodeum rugose; tergites 1 and 2 shiny, tergite longitudinally striate;
	clypeus about 3× as long as wide Baeognatha
45 (30).	Spiracle on tergite 1 at or behind middle (except in Euphoriana, but radius
	ends before apex of wing); abdomen petiolate or subpetiolate; radius usual-
	ly ending before apex of wing (figs. 43-47) Euphorinae, 46
	Spiracle on tergite 1 definitely before middle; abdomen sessile; radius usually
	ending at or near apex of wing 54
	EUPHORINAE
46 (45).	Tergite 1 petiolate, slender and subcylindrical; tergite 2 laterally smooth and
	convex with dorsal part, its epipleura absent; ovipositor short
	Tergite 1 subpetiolate, broadening strongly behind middle and depressed;
	tergite 2 with lateral edge sharp, epipleura present (except in Syntretus) 48
47 (46).	Fore wing with 3 cubital cells; medius distinct in its entire length (fig. 43);
	frons with median carina; antenna always with 18 segments; face broad,
	about $2 \times$ as wide as high Aridelus Fore wing with 2 cubital cells; medius basally weak (fig. 45); frons without
	median carina; antenna with more than 18 segments; face subquadrate
	Wesmaelia
48 (46)	Fore wing with 3 cubital cells (fig. 42) Meteorus
40 (10).	Fore wing with 2 cubital cells, rarely 1
49 (48).	First cubital and first discoidal cells separate
	First cubital and first discoidal cells confluent
50 (49).	Medius well developed in its entire length (fig. 44) Perilitus
	Medius obsolete or weak basally (fig. 47)
51 (50).	Cubitus, intercubitus, discoideus and recurrent veins effaced Euphoriana
	Cubitus distinct at least at base; first cubital and first discoidal cells distinct-
	ly separate; recurrent vein rarely absent (fig. 47) Euphorus

52 (49).	Scape	unusally	long, a	t least	1/2 as	long	as height	of he	ad (fig. 24) Streblocera
	Scape	very sho	rt, neve	er more	than 2	$2 \times as$	long as	thick		53

- 53 (52). Medius and nervellus well developed (fig. 46); tarsal claws simple ... Microctonus Medius and nervellus weak or effaced (fig. 48); tarsal claws bidentate... Syntretus

CARDIOCHILINAE

55 (54). Hind basitarsus unusually broadened and lengthened; propodeum mo	ore or
less triangular from dorsal view La	ıminitarsus
Hind basitarsus normal, not as above; propodeum more or less semici	rcular
from dorsal view C	ardiochiles
56 (54). Occipital carina absent Macrocentrinae, Ma	acrocentrus
Occipital carina entire or dorsally interrupted	57
57 (56). Fore wing with 2 cubital cells	acinae, 58

BLACINAE

(excl. Stantonia, Urosigalphus and Triaspis)

- 62 (61). Anal cell of fore wing without a cross-vein (fig. 53); head usually transverse, sometimes cubical; frons not or slightly depressed....... Opiinae, Opius Anal cell of fore wing with 1 or 2 interanals (figs. 54-56); head usually conical or subcubical; frontal depression more or less deep Helconinae, 63

HELCONINAE

	Cubital cell 2 rhombic (fig. 55)
65 (64).	Abdomen attached high on propodeum, distant from hind coxa Cenocoelius
	Abdomen attached low on propodeum, close to hind coxa
66 (65).	Hind femur enlarged and with strong ventral tooth and or a row of blunt
	spines; interanella present Helconidea
	Hind femur slender and without ventral spines; interanella absent Helcon
67 (64).	Clypeus with median apical tooth (fig. 25); scape about $3 \times$ as long as wide;
	clypeal fovea nearer to the other fovea than to malar groove Baeacis
	Clypeus without median apical tooth; scape about $2 \times$ as long as wide; cly-
	peal fovea equidistant to other fovea and malar groove Diospilus
68 (18).	Occipital carina present; mesopleural furrow and prepectal carina present;
	submediellan variable in size
	Occipital carina absent; mesopleural furrow and prepectal carina absent ex-
	cepting the genera included in couplets 110 and 111; submediellan small 101
69 (68).	Fore wing with 2 cubital cells (intercubitus 2 absent) (figs. 57 & 58)
	Hecabolini, 70
	Fore wing with 3 cubital cells (fig. 59)
	HECABOLINI
70 (69).	Submediellan cell absent; fore wing with discoidal cell 2 closed (fig. 57);
	tergite 1 longitudinally striate; \mathcal{J} and \mathcal{P} dimorphic, \mathcal{J} with tergites beyond
	the second unusually elongate and depressed, ♀ abdomen normal Aivalykus
	Submediellan cell present; fore wing with discoidal cell 2 open (fig. 58);
	tergite 1 variable in sculpture; \Im and \Im similar
71 (70).	Thorax and propodeum elongate and depressed; tergite 2 with a V-shaped
	groove or carina; scape about $2 \times$ as long as pedicel; medium-sized species
	Thorax and propodeum normal, not unusually elongate nor depressed; tergite
	2 without V-shaped groove or carina; scape subequal to pedicel; small
	species Monolexis
72 (69).	Abdomen petiolate, tergite 1 not gradually widened apically; epipleura not
	reaching middle of tergite; suture between tergites 2 and 3 absent (but
	faintly impressed in Pseudospathius) True Spathiinae, 73
	Abdomen subpetiolate or sessile; epipleura reaching middle of tergite; suture
	between tergites 2 and 3 present
	SPATHIINAE
73 (72).	Epipleura of abdominal segments 2 and 3 separated by a suture; species
	large, about 12 mm long excluding ovipositor; head and thorax yellowish,
	abdomen entirely black Pseudospathius
	Epipleura of abdominal segments 2 and 3 confluent; species less than 12 mm
	long; almost entirely black species74
74 (73).	Scape elongate and slender, about $2 \times$ as long as wide; hind coxa subequal
	in length to the swollen hind femur, the latter abruptly narrowed at base;
	species slender, gaster elongate Paraspathius
	Scape short and cup-shaped, usually not more than $1.5 \times$ as long as wide, if
	somewhat longer, anterior face of mesoscutum perpendicular; hind coxa

without basal spine on inner side; pronotal collar not swollen... Rogadinae, 76 Ovipositor long, longer than greatest depth of abdomen; head usually cubical; fore tibia usually armed with row of short spines on inner side; hind coxa with or without basal spine on inner side; pronotal collar usually swollen... 92

ROGADINAE

76 (75). Propodeum areolated, at least 1 closed area present; species small, about 77 (76). Only tergites 1-3 visible above, longitudinally striate throughout; suture between tergites 2 and 3 deep 78 All tergites visible above, not longitudinally striate throughout or striate on 78 (77). Apical margin of tergite 3 transparent and notched at middle..... Aulosaphes Apical margin of tergite 3 with a strong tooth on each corner...... Acanthormius 79 (77). Propodeum with 2 strong teeth; suture between tergites 2 and 3 absent; recurrent vein interstitial with intercubitus 1..... Pambolus Propodeum without teeth; suture between tergites 2 and 3 present; recurrent 80 (76). Legs with last tarsal segment enlarged, tarsal segments 2, 3, and 4 short and wide; claws pectinate; femora short and thick...... Yelicones Legs with last tarsal segment not enlarged; tarsal segments 2, 3, and 4 long and slender; claws simple with or without a basal lobe; femora slender..... 81 81 (80). Mid and hind tibial spurs strongly curved, long and bare or hairy on basal Mid and hind tibial spurs straight, short and hairy throughout; postnervellus 82 (81). Tarsal claws without basal lobe; propodeal spiracle small and circular; tergite 1 at most $2 \times$ as long as apical width; maxillary palpi of \mathcal{F} enlarged and flattened, of \mathcal{P} slender..... Dedanima Tarsal claws each with a basal lobe; propodeal spiracle usually elliptical; tergite 1 is $2-3 \times$ as long as apical width; maxillary palpi of 3 not enlarg-83 (82). Intercubitus 1 and abscissa 2 of radius equal in thickness, intercubitus 1 straight and oblique; maxillary palpi of \mathcal{J} enlarged, in some species labial palpi enlarged also; in the Philippine species, tergite 1 is $2 \times$ as long as apical width Macrostomion Intercubitus 1 and abscissa 2 of radius thickened at the junction and together forming a curve; maxillary palpi of \mathcal{J} slender or unmodified except in 1 Philippine species (*Megarhogas stigmaticus* Baker, \mathcal{F} heretofore not known); tergite 1 is 2.5-3× as long as apical width Megarhogas

Baltazar: Parasitic Hymenoptera in the Philippines

767

84 (81). Postnervellus present (fig. 61)						
Postnervellus absent (fig. 60) 86						
85 (84). Tergites beyond third retracted underneath; hind tarsal spurs wide apart						
basally Clinocentrus						
Tergites all visible dorsally; hind tarsal spurs close together basally						
A few species of Rogas						
86 (84). Propodeal teeth present or with a slight suggestion; tarsal claws each with						
basal lobe						
Propodeal teeth absent; tarsal claws without a basal lobe						
87 (86). Tergite 1 sessile; propodeal teeth strong; middle lobe of mesoscutum promi-						
nently bulging and higher than lateral lobes Conspinaria						
Tergite 1 subpetiolate; propodeal teeth weak; middle lobe of mesoscutum not						
prominent, sloping anteriorly and not higher than lateral lobes Gyroneuronella						
88 (86). Notaulus weak Hemigyroneuron						
Notaulus deeply impressed 89						
89 (88). Cubital cell 2 smaller than cubital cell 3, less than $1/2$ size of 3; nervulus						
strongly postfurcal, distance from basal vein usually equal to or greater						
than its length; nervellus straight (fig. 61) Most species of Rogas						
Cubital cells 2 and 3 subequal in size; nervulus slightly postfurcal or inter-						
stitial, its distance from basal vein less than $1/2$ its length; nervellus curved						
(fig. 60)						
90 (89). Tergites 4-6 shorter than length of 2 and 3 united Pelecystoma						
Tergites 4-6 equal to length of 2 and 3 united						
91 (90). Nervulus interstitial Rhogasella						
Nervulus slightly postfurcal (fig. 60) Pseudogyroneuron						
92 (75). Tergite 1 sessile, if petiole is elongate the rest of tergites longitudinally striate;						
petiole usually longitudinally striateDoryctinae, 93						
Tergite 1 subpetiolate, widened at apex, 2.5 or more times as long as apical						
width; petiole coarsely punctate or transversely striate but not longitudinal-						
ly striate						
DORYCTINAE						

STEPHANISCINAE

- 99 (98). Postnervellus curved outwards (fig. 64); tergite 1 very long, about 6× as long as its apical width, with fine transverse aciculae......Leptospathius Postnervellus straight; tergite 1 long, about 3× as long as its apical width, rugoso-punctate......Halycaea
 100 (98). Fore wing speckled with fuscous and white; fore and mid femora with dorsal blister-like swelling near base; pronotal collar gradually sloping anteriorly, not swollen......Platyspathius

BRACONINAE

101 (68). Prepectal carina (fig. 21) and mesopleural furrow present; nervulus post-
furcal (fig. 65) 102
Prepectal carina and mesopleural furrow absent; nervulus variable in posi-
tion, usually interstitial 104
102 (101). Pronotum without dorsal spine; tergite 5 with 4 apical spines; propodeum
without teeth; tarsal claw without a basal lobe Batotheca
Pronotum with a dorsal spine; tergite 5 with 1 median apical spine or none;
propodeum with a pair of teeth; tarsal claw with a basal lobe 103
103 (102). Tergites 3-5 without spines; pronotal spine short and straight; hind femur
with a preapical ventral tooth; propodeal teeth not distinct Spinariella
Tergites 3-5 with spines; pronotal spines long and curved forward; hind
femur without a tooth; propodeal teeth distinct (fig. 21) Spinaria
104 (101). Abdomen carapace-like with 2 apical spines
Abdomen not as above 105
105 (104). Nervulus antefurcal; abscissa 1 of cubitus strongly curved (fig. 70); dis-
coideus thickened; eyes large and deeply emarginate Aphrastobracon
Nervulus interstitial or postfurcal; abscissa 1 of cubitus not as above; dis-
coideus not thickened; eyes variable in size
106 (105). Scape excised basally (fig. 25) 107
Scape not excised basally108
107 (106). Notaulus absent; thorax, propodeum and tergites depressed; pronotum pro-
longed into a neck Chaoilta
Notaulus present; thorax, propodeum and tergites normal; pronotum not
prolonged into a neckAtanycolus

108 (106). Tergite 1 is $1.5-3.0 \times$ as long as its apical width; tergite 2 is 0.7-1.5 as long as its apical width; nervulus not forming straight line with basal vein,
the latter slanting and forming a 45° -60° angle with subcosta (figs. 66-68);
head usually cubical; mostly large species
Tergite 1 shorter than or at most as long as its apical width, or with a
wide membrane on its side; tergite 2 short, 0.4–0.5 as long as its apical
width; nervulus usually forming a straight line with basal vein, the latter
forming a 75° -80° angle with subcosta (figs. 71-73); head usually trans-
verse; mostly medium-sized to small species
109 (108). Notaulus deeply impressed on entire length of mesoscutum; tergite 2 long,
$1.2-1.5$ as long as its apical width; subgenital plate of φ pointed apical-
ly and extending beyond tip of last tergite $\dots \dots \dots$
Notaulus shallow and impressed only on anterior 0.4 of mesoscutum; tergite
2 as long as or shorter than its apical width (excepting 3^{3} of <i>Euuro</i> -
bracon); subgenital plate of φ variable in shape and length
110 (109). Radial vein ending before apex of wing (fig. 68); scape short, as long as
its diameter; tergite 2 without a midbasal triangular area nor carinae;
wings mottled with fuscous
Radial vein ending at apex of wing; scape longer than its diameter; tergite
2 with a wrinkled midbasal triangular area and 2 apically convergent lat-
eral carinae; wings banded yellow basally and black apically Gronaulax 111 (109). Tergite 2 without midbasal triangular area; scape short, $1.0-1.5 \times$ as long
as its diameter
Tergite 2 with midbasal triangular area; scape usually long, $2.0-3.0 \times$ as
long as its diameter
112 (111). Recurrent vein strongly antefurcal, its distance from intercubitus 1 equals
0.5-1.0 the length of abscissa 1 of radius (fig. 66); tergite 3 without tri-
angular area on each basal corner; nervulus postfurcal; tergite 1 with
deep median groove on basal 0.3 Euurobracon
Recurrent vein interstitial or slightly antefurcal (fig. 67); tergite 3 with tri-
angular area on each basal corner; nervulus almost intestitial or postfur-
cal; tergite 1 without deep median groove basally
113 (112). Radius ending before apex of wing Vipio
Radius ending at apex of wing114
114 (113). Nervulus postfurcal; ovipositor long and auger-like, with 4 constrictions at
apexBathyaulax
Nervulus interstitial; ovipositor short, thickened and curved downwards
Iphiaulax
115 (111). Tergite 2 without pair of convergent carinae, no tooth on each apical cor-
ner; scape short, 1.0-1.5 as long as its diameter Iphiaulax
Tergite 2 with pair of convergent carinae apically and with small tooth on
each apical corner; scape long, $2-3 \times$ as long as its diameter116
116 (115). Tergite 1 with median carina; scape long, length about $3 \times$ diameter
Cratobracon
Tergite 1 without median carina; scape shorter, length about $2.0 \times$ diame-
ter Sigalphogastra

1962

~

117 (108).	Recurrent vein strongly antefurcal, its distance from intercubitus 1 is 0.5– 1.2 as long as abscissa 1 of radius (fig. 69); tergite 2 without median tri- angular area basally
	Recurrent vein interstitial or slightly antefurcal (figs. 72 & 73); tergite 2
	with or without median triangular area basally
118 (117).	Tarsal claws without basal lobe
	Tarsal claws each with basal lobe 119
119 (118).	Middle lobe of mesoscutum without longitudinal grooves; tergite 2 with 2 grooves convergent apically; cubital cell 2 small (fig. 69); last tergite
	with apical margin convex Tropobracon
	Middle lobe of mesoscutum with 2 longitudinal grooves, tergite 2 without
	grooves; cubital cell 2 variable in size; last tergite with its apical mar-
	gin notched medially or with apical spines 120
120 (119).	Tergite 6 with 3 apical spines; cubital cell 2 small as in Tropobracon
	Odontopygia
	Tergite 6 with median apical notch; cubital cell 2 large Dioxybracon
121 (117).	Tergite 1 flat and long, about $3 \times$ as long as its apical width, with wide membrane laterally; all tergites smooth and shiny Myosoma
	Tergite 1 short, length equal to apical width or less, lateral membrane ab-
	sent; tergites sculptured variably
122 (121).	Apical margin of tergite 5 with numerous teethOdontogaster
	Apical margin of tergite 5 without teeth123
123 (122).	Suture between tergites 2 and 3 deep and sinuate 124
. ,	Suture between tergites 2 and 3 straight or slightly sinuate 126
124 (123).	Antennal socket raised and toothed dorsally; eyes bare Hemiglyptus
	Antennal socket not raised and without tooth; eyes with very short hairs 125
125 (124).	Hind tibia noticeably broadened and flattened; cubital cell 2 subequal to
	3 in length; tergites 4-7 gradually becoming shorter, visible dorsally
	Cratocnema
	Hind tibia not as above; cubital cell 2 about $1/2$ as long as 3; tergites 4-
	7 short and telescoped Philomacroploea
126 (123).	Abscissa 1 of cubitus curved at base (fig. 72); cubital cell 2 subequal to 3
	in length; tergites 3-5 each with a transverse groove along apical mar-
	gin, recurrent vein usually interstitial Campyloneurus
	Abscissa 1 of cubitus straight (fig. 73); cubital cell 2 equal to or shorter
	than 3; tergites 3-5 usually without transverse groove apically; recurrent
	vein antefurcal or interstitial Bracon

LITERATURE CITED

		Press/ 1	r outuroga	01	1 mmppme	Hymenoptera,	WITCH W	olonography
(1758	-1958).	Philip. J	our. Agric.					

- 1962. Ettchellsia philippinensis, new species (Megalyridae, Hymenoptera). Philip. Jour. Sci. 90 (2): 219-20.
- Brues, C. T., A. L. Melander & F. M. Carpenter. 1954. Classification of insects. Mus. Comp. Zool., Bull. 108: 626-52.

- Krombein, K. V. et al. 1958. Hymenoptera of America North of Mexico, Synoptic Catalog. U. S. Dept. Agric. Monograph No. 2, First Supplement: 1-100.
- Maa, T. C. & C. M. Yoshimoto. 1961. Loboscelidiidae, a new family of Hymenoptera. Pacific Insects 3 (4): 523-48.
- Muesebeck, C. F. W. 1936. The genera of parasitic wasps of the braconid subfamily Euphorinae, with a review of the nearctic species. U. S. Dept. Agric. Misc. Publ. No. 241: 1-37.

et al. 1951. Hymenoptera of America North of Mexico, Synoptic Catalog. U. S. Dept. Agric. Monograph No. 2: 89-734.

- Nixon, G. E. J. 1943. A revision of the Spathiinae of the Old World (Hymenoptera, Braconidae). Roy. Ent. Soc. Lond., Trans. 93: 173-456, 1 pl.
- Pierce, W. D. 1928. Notes on the sugar cane insect situation in Negros. Sugar News (Manila) 9: 616–25.
- Richards, O. W. 1956. Hymenoptera-Introduction and keys to families. Handbooks for the identification of British insects. London: Roy. Ent. Soc. Lond. 6: 1-94.
- Rohwer, S. A. & A. B. Gahan, 1916. Horismology of the hymenopterous wing. Ent. Soc. Wash., Proc. 18: 20-76.
- Townes, H. 1948. Key to the families of the commoner Clistogastra. IN Comstock's An Introduction to Entomology. New York: Comstock Publ. Co., 9th ed., 892–94.
 - ------ 1950. The nearctic species of Gasteruptiidae (Hymenoptera). U. S. Nat. Mus., Proc. 100: 85-145.

1956. The nearctic species of trigonalid wasps. Ibid. 106: 295-304.

- Weld, L. H. 1952. Cynipoidea (Hym.) 1905–1950. Privately printed Ann Arbor, Michigan, 351 pp., illus.
- Williams, F. X. 1928. Studies in tropical wasps.—their hosts and associates (with descriptions of new species). Hawaiian Sugar Plant. Assn. Ent. Ser., Bull. 19: 19-20.
- Yasumatsu, K. & H. Kuroko. 1957. The occurrence of Mesocoelus philippinensis Muesebeck in Japan (Hym., Braconidae). Sci. Bull. Fac. Agric., Kyushu Univ. 16: 221-24.